

Microplastic Management and Cardiovascular Risk: Mitigating the Threat of Plastic Contaminants in Atherosclerotic Plaque

Dr. USHAA ESWARAN¹, PRADEEPA K², R. THIYAGARAJAN³, K. RAMYA SREE⁴, UMA MAHESWARI P⁵

Mahalakshmi Tech Campus, Chennai, Chrompet-600044

*Corresponding Author

Article History

Received: 07.10.2025

Revised: 29.10.2025

Accepted: 12.11.2025

Published: 01.12.2025

Abstract:

PL pollution is caused by contemporary society's unsustainable consumption and dumping of PL items, placing economies, landscapes, and people's health in risk. So far, remediation operations have attempted to mitigate the negative effects of PL pollution, but efforts are still unable to cope with the increased volumes of PL released to the environment. As a consequence, emphasizing a standardized measurement strategy to recycle PL emissions into the environment is essential. Badly run garbage is a significant ground cause of PL pollution that could be reduced by changing life of PLs, especially in terms of production, usage, and destruction, with an Effective Waste Management System. In this review article, we describe existing techniques for improving the life span and waste disposal of PLs that may be applied to decrease PL waste and environmental and health implications. (1) Regulatory oversight of manufacturing and utilization; (2) eco-design; (3) rising the demand for recycled PLs; (4) lowering the use of PL products; (5) use of renewable power for reprocessing; (6) extended producer responsibility over waste; (7) advancements in collection systems; (8) good planning of composting; (9) utilisation organic and degradable PL products; and (10) advancement in recyclability are among the ten suggestions for decision makers to reduce its environmental footprint.

Keywords: Microplastic, microplastic conversion, source, types and health impacts.

INTRODUCTION

"Pliable and readily made" was the original definition of plastic (PL). It wasn't until recently that the name "polymer" was used for a group of materials. Polymers (PLs) are long chains of molecules with the word "polymer" meaning "many parts." In nature, PLs abound. A common natural polymer is cellulose (CL), which is found in cellular membranes. During the preceding century and a half, humans figured out how to make synthetic PLs, sometimes using natural materials like CL, but more often with the copious carbon atoms provided by petroleum-based goods. Synthetic PLs are made up of lengthy chains of atoms arranged in repeating units, which are often much longer than natural PLs. Because of the length of the chains and the patterns in which they are arranged, PLs are strong, lightweight, and elastic. To put it another way, that's what gives them their flexibility. Such characteristics that make synthetic PLs incredibly beneficial, and we've been using them since we learned how to produce and control them. Especially in last 50 years, PL have penetrated our culture and influenced how we live [1].

First Synthetic PL

In answer to a \$10,000 reward offered by a New York corporation for anybody who could make an ivory substitute, John Wesley Hyatt created the first synthetic PLs in 1869. Billiards' popularization had put a huge strain on natural ivory resources, that were acquired by killing wild elephants. By mixing cotton CL with camphor, Hyatt found a PL that can also be shaped into a range of shapes and molded to simulate natural substances such as tortoiseshell, horns, linen, and ivory.

This discovery was revolutionary. Human output was not bound by natural restrictions for the very first period in history. Mother Earth could only provide far more timber, metal, rock, bones, ivory, and horn. Humanity, on the other hand, are concerned with producing substances [1]. Not only did this technology assist people, but it also helped the ecosystem. In commercials, celluloid was hailed as the saviour of the elephant and tortoise. PLs have the ability to preserve the ecological landscape from the detrimental effects of human demand. The invention of new substances also liberated humans from the economic and social constraints imposed by a shortage of mineral wealth. Due to the obvious inexpensive cost of celluloid, material luxury has become more fashionable and attainable. As well as the PL revolutionary was only getting underway [2].

Development of New PLs

Leo Baekeland invented Bakelite, a first completely synthetic polymer, in 1907. It didn't include any molecules found in nature. Baekeland had been seeking for a synthetic substitute to shellac, a natural electrical insulator, to satisfy the demands of the quickly electrifying United States. Bakelite was not only a good insulator, but it was also lengthy, high - temperature, and, like celluloid, well with the mass production. "The material with a thousand uses" is how the chemical is promoted. Bakelite could be moulded or moulded into whatever, opening world of possibilities. Major chemical businesses were inspired by Hyatt and Baekeland's success to invest in the research and innovation of additional PLs, as well as other PLs swiftly joined celluloid and Bakelite. Whereas Hyatt and Baekeland are seeking for substances with certain

characteristics, the latest research initiatives are seeking for novel PLs in the hopes of finding future applications for them [1,2].

Age of PL

World War II (WWII) necessitated a substantial expansion of the PLs sector in the US, as economic growth was as important as military victory. The need to conserve precious natural resources spurred the development of synthetic fibers to the top of the priority list. Options were provided by PLs. Nylon was invented by Wallace Carothers in 1935 as a synthetic silk, and it was widely used in the war for parachutes, ropes, body armour, helmet liners, and other products. Areophane windows were replaced with Plexiglas. As a consequence of the fight, "PLs have indeed been turned to specific uses, and the adaptability of PLs has been proved all over again," as shown in a Time magazine report. During WWII, PL output in the United States rose by 300 percent [1]. After the war, the increase in PL production continued. Americans were ready to spend again after the Great Depression and WWII, and the majority of what they bought was made of PL. As per author Susan Freinkel, "PLs challenged traditional substances and won in product after product, market after market, filling the role of steel in automobiles, paper and glass in containers, and timber in furniture." The promise of PLs gave some onlookers a near-utopian vision of a future filled with great material riches thanks to an inexpensive, safe, and sanitary material that individuals could shape with their every want [2].

Growing concerns about PLs

The perfect hope of PL was short-lived. In the postwar years, PLs were no longer seen as unquestionably good, leading to a shift in American attitudes. The discovery of PL garbage in the seas occurred in the 1960s, during which time Americans were increasingly aware of environmental concerns. Rachel Carson's book *Silent Spring*, published in 1962, highlighted the dangers of pesticides. In 1969, there was a large oil spill off the coast of California, and the polluted Cuyahoga River in Ohio caught fire, raising environmental concerns. Observers were concerned about the permanency of PL rubbish as public awareness of environmental problems expanded. PL started to be used to denote anything inexpensive, flimsy, or deceptive over time. An elder acquaintance pushed Dustin Hoffman's character in *The Graduate*, one of the finest films of 1968, to seek a career in plastics. Audiences grumbled at what they saw as misguided enthusiasm for a business that, rather than being full of potential, was a symbol of cheap uniformity and artificiality, as Hoffman did [2].

Effect of PL on human health

As public concern about waste developed in the late 1970s and early 1980s, PL's reputation suffered dramatically. Because, whereas many PL things are

disposable, PL persists in the environment permanently, it became a special emphasis. The PLs industry offered recycling as a solution. The plastics sector led a successful push in the 1980s to persuade municipalities to collect and process recyclable materials as part of their waste-management systems. Recycling, on the other hand, is far from perfect, and the great majority of PLs still end up in landfills or the environment. PL supermarket bags have been a target for environmentalists aiming to limit single-use, throwaway PLs, and bag bans have already been passed in a number of places across the United States. The Great Pacific Debris Patch, a swirl of PL debris the size of Texas drifting in the Pacific Ocean, is the epitome of the PL waste dilemma. As a result of growing concern about the potential harm they pose to human health, PL's image has deteriorated further. The chemicals, such as phthalates, that are introduced into plastics throughout the manufacturing process to make them more flexible, durable, and transparent are at the focus of these issues. Some researchers and members of the public are concerned that these chemicals are leaking from PLs and making their way into our food, water, and bodies. The endocrine system can be influenced by these drugs in exceedingly high concentrations. Researchers are especially worried about the effects of these chemicals on children, as well as the long-term consequences of their accumulation for future generations [1,2].

PLs future

Despite growing scepticism, PLs are critical in today's world. Computers, mobile phones, and the vast majority of life-saving advances in modern medicine were all made possible by PLs. PLs are light and insulating, reducing the usage of fossil fuels for heating and transportation. Low-cost plastics, maybe most crucially, raised people's living standards and increased access to material goods. If PLs were not accessible, many of the items we take for granted would be unavailable to everyone except the wealthiest Americans. Many of our products have been made cheaper, lighter, safer, and stronger by substituting PL for natural materials. Since PLs are clearly crucial in our life, several scientists are striving to make them safer and more long-lasting. Certain businesses are developing bioplastics, which are made from plant crops rather of fossil fuels, to make chemicals that are more environmentally friendly than standard PLs. Others are working to create PLYs that are completely biodegradable. One of the ambitions of some innovators striving to enhance recycling efficiency is to develop a technology that converts PLs back into the fossil fuels from which they originated. According to all of these creators, PLs aren't perfect, but they're an important and critical part of our future [1,2].

PL to microplastic (MP) conversion

MPs are microscopic PL particles with a diameter of less than 5 millimetres. In water, MPs are

invisible and float or sink depending on their composition. Because MPs like polypropylene are lighter than seawater, they float and disperse widely over rivers. Acrylic and other MPs have a higher density than seawater and will collect in the ocean's deepest depths. About 99 percent of the PL in the ocean is thought to be made by MPs. Floating MPs will eventually assemble in massive gyres. MPs that sink seeking food are perplexed by sea life. The hadal zone (the deepest part of the ocean) may be one of the most important MPs sinks on the planet. Microfibers (MFs) are included in MPs. They include microscopic threads that enter the water when synthetic clothing like polyester and nylon is washed [3-5].

Sources of MP pollution

Automobile industry: Tyre wear and tyre dust (Adachi and Tainosho, 2004), emissions from non-exhaust vehicles [6, 7], road dust resuspension [8], leachate from weathering PLs [9], brake friction materials [10], brake pad materials [11], pavement-tire interface ultrafine particles [12], tire tread wear particles [13] and lithium-ion battery packs [14].

Textile industry: Polyester clothing [15], MFs produced by washing dryers [16], 3-D printing [17] and synthetic fibers from textiles [18].

Food industry: PLY film packaging for food [19], PL-coated paper products [20], PL bags [21], PL straws [22] and disposable PLs and wooden chopsticks [23].

Cosmetic industry: Facial cleansers [24], microbeads (MBs) in facial scrubs [25, 26] and hair conditioner and eye shadow [27].

Health industry: Nylon flock [28-30], COVID-19 pandemic face mask [31], medical waste disposal [32] and PVC medical product waste [33].

Fisheries and aquaculture: Marine paints (antifouling agent) [34], resin pellets [35], bisphenol A [36], derelict traps [37], polystyrene spheres [38], PET bottles [39], phthalate esters [40], polyethylene wear debris [41], 4-nonylphenol [42], HDPE cages [43], Bisphenol S [44], Polybrominated diphenyl ethers [45], polyfilament nylon fragments [46], pelagic plastic and tar [47], neuston plastic [48], polychlorinated biphenyls [49], fibrinogen-coated polystyrene [50], LDPE [51], 60 nm polystyrene particles [52], polystyrene microspheres [53], Styrofoam debris [54], styrene oligomers [55], plastic garbage that floats [56], polyethylene microbeads [57], spirocyclic polyacetal ethers [58], nylon threads [59], Hexabromocyclododecane [60], dioxin-like PCBs [61], polypropylene [62], polyamide 6 multifilament fishing net [63], Polystyrene nanoplastic [64], nonylphenol [65] and micro-sized PVC particles [66].

RESULTS AND OBSERVATIONS:

Types of MPs

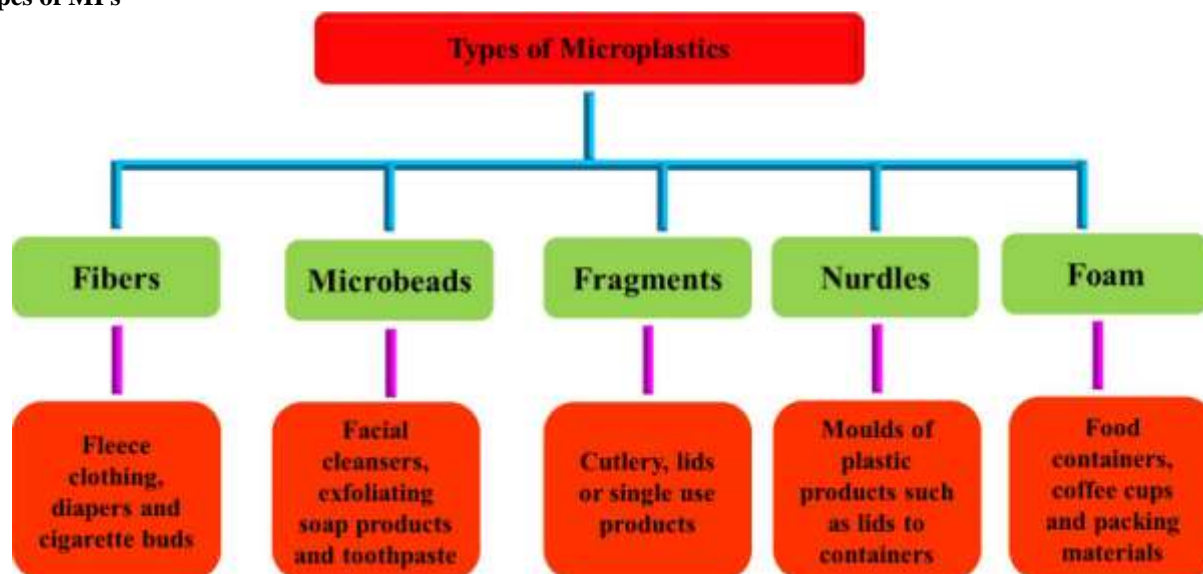


Fig. 1: Five types of MPs

MPs are divided into two types: primary and secondary. Primary MPs include MBs in personal care products, PL pellets or nurdles used in industrial manufacturing, and PL fibres used in synthetic textiles (e.g., nylon). Primary MPs enter the environment through product consumption (e.g., personal care goods rinsed into wastewater systems from homes), unintentional loss from spills during manufacturing or transportation, or abrasion during washing (e.g., laundering of clothing made with synthetic textiles). When larger polymers deteriorate due to weathering, such as exposure to wave action, wind abrasion, and UV radiation from the sun, secondary MPs emerge (Fig. 1) [67].

Diseases caused by MPs

Human beings: Crohn's disease [68], Endoplasmic reticulum stress [69], endocrine-disrupting substances' potential for causing harm to humans [70], Oral squamous cells carcinoma [71], MPs absorption in human stool [72], clathrin-mediated endocytosis [73], gastric adenocarcinoma [74], mitochondrial injury pathways [75], metabolic disorders in the offspring [76], immunotoxicity [77], disruption of thyroid hormone [78], Coronary Artery Disease (CAD) [79,80] and oxidative stress [81].

Aquatic animals: Inflammation of the intestine, oxidative stress, vascularization, and metabolome and microbiome abnormalities in zebra fish [82, 83], *Pomatoschistus microps* predatory performance and efficiency are reduced [84], hemocytopenia in crab [85], hepatic stress in the fishes [86], toxicity in Antarctic Krill [87] and endocrine disruption in adult fish [88].

Birds: Chemicals generated from PL accumulate in marine birds [89], endocrine disturbance and reproductive toxicity [90], MP ingestion and dispersion by vultures [91], cerebellar toxicity in Quails [92] and kidney injury in Quails [93].

Ruminants: Sheep and goats' digestive tracts accumulate PL detritus [94].

Techniques used to detect, identify and quantify MPs

ICP-MS [95], micro-ATR-FTIR [96], TEM and GC-MS [97], TGA-FTIR-GC-MS [98], Pyrolysis-GC-MS and FTIR [99], FTIR imaging [100], TGA/DSC-FTIR [101], DSC [102], TGA-DSC [103] and AFM-IR [104].

Methods used to degrade MPs

A modern strategy to developing novel synthetic PLs is based on their life span and takes into account environmental effect from conversion to disposal [105]. Polymeric materials endanger natural habitats and the quality of life on the earth when improperly disposed or under unmanaged situations [106]. Few products are designed with the end user in mind (management of waste or reprocessing) in mind, incredibly simple PLs have been extensively denounced for their visual pollution, difficulties in eliminating them from the ecosystem, and great resilience to deterioration.

Degradation is a synthetic process that causes the splitting of PL chains and can irreversibly change the characteristics of PL composites. A number of physical and chemical variables can cause it to occur [107]. One of the decade's greatest issues is determining how long a material can withstand prior to actually deteriorating or anaerobic decomposition, so that it could be disposed of in an environmentally friendly manner. According to research, thermoplastic materials degrade after being dumped in a number of ways, which might happen slowly or quickly depending on the environment and molecular makeup. Unlike biodegradable PLs, which can include heteroatoms in the molecular chain and so disintegrate quickly when exposed to the right conditions, saturated polymeric chains do not allow for microbial degradation [108].

Gradually degradable or non-biodegradable PLs have been studied as a greener alternative to biodegradable PLs like chitosan or CL. However, it is widely acknowledged that today's biodegradable PLs have unsatisfactory mechanical or surface properties for some applications, and that some of them are costly, restricting their use in select applications [109]. The density of PLs in compared to the density of saltwater influences whether they float or sink when they reach the marine environment. The type of a PL and the conditions to which it is exposed, which might range from abiotic to microbial absorption, impact its degradation process [110]. As a result, either abiotic or biotic degradation of PLs may be classified. The damage caused by environmental variables such as temperature, UV irradiation, wind, and waves is referred to as abiotic deterioration. Biotic degradation, on the other hand, is characterized as biodegradation caused by microorganisms (MOs) that alter and consume the PL, altering its properties (Table 1). Both forms of deterioration are often active at the same time in nature [111].

Table 1: List of MOs used for biodegradation of PL and MPs

Name of the MOs	PL/MPs	References
<i>Bacillus subtilis</i>	PET	[112]
<i>E. coli</i>	PET	[113]
<i>Ideonella sakaiensis</i>	PET	[114]
<i>Ideonella sakaiensis</i>	PET	[64]
<i>Ideonella sakaiensis</i>	PET	[115]
<i>Spirulina species</i>	PET	[116]
<i>Listeria monocytogenes</i>	PET	[117]
<i>Penicillus simplicissimus</i>	PET	[118]
<i>Brevibacillus borstelensis</i>	PET	[119]
<i>Streptomyces species</i>	PET	[120]
<i>Streptomyces species</i>	Poly (3-hydroxybutyrate) and poly (3-hydroxybutyrate-co-3-hydroxyvalerate)	[121]
<i>Alcaligenes faecalis</i>	Poly (3-hydroxybutyrate)	[122]
<i>Alcaligenes faecalis</i>	Polycaprolactone	[123]

<i>Comamonas acidovorans</i>	Polyester polyurethane	[124]
<i>Phanerochaete chrysosporium</i>	Polyvinyl chloride	[125]
<i>Thermomonospora fusa</i> and <i>Fusarium solani</i>	PET	[126]
<i>Comamonas acidovorans</i>	Polyurethane	[127]
<i>Pseudomonas putida</i>	Phenylacetic acid	[128]
<i>Pseudomonas</i> and <i>Bacillus</i> species	PET	[129]
<i>Pseudomonas putida</i>	PET	[130]
<i>Pseudomonas aeruginosa</i> and <i>Achromobacter species</i>	Polyvinyl chloride	[131]
<i>Acinetobacter species</i>	PET	[132]

Biodegradation of MP using algae

In sewage water, algae have been discovered to grow on artificial substrata such as polythene surfaces, and these colonizing algae have been proven to be less harmful and non-toxic [133]. The biodegradation of PLY begins with algae adhering to the surface, and their production of ligninolytic and exopolysaccharide enzymes is essential [134]. Algal enzymes in the liquid media interact with macromolecules on the PL surface, causing biodegradation to occur [135]. Algae use PLY as a carbon source because species that grow on the polyethylene surface have larger cellular contents and a faster specific growth rate [134]. Furthermore, the transverse section of the algal-colonized polyethylene sheets has clearly shown surface deterioration or disintegration [136]. Fouling, corrosion, hydrolysis and penetration, breakdown of leaching components, and pigment colouring via diffusion into the PLY are all examples of biodegradation techniques that have been documented in previous studies. *Anabaena spiroides*, a blue-green alga, degraded the lowest density polyethylene, followed by the diatom and the green alga [136]. Sarmah and Rout (2018) [134] found that readily available, fast-growing, and easily isolable freshwater nontoxic cyanobacteria are capable of colonizing the polyethylene surface and biodegrading low-density polyethylene efficiently without any pretreatment or pro-oxidant additives (Table 2) [137].

Table 2: List of algae used for biodegradation of PL and MPs

Name of the algae	PL/MPs	References
<i>Uronema africanum</i>	LDPE	[138]
<i>Chlorella</i> and <i>Cyanobacteria species</i>	LDPE	[139]
<i>Sargassum linifolium</i>	PET	[140]
<i>Chlamydomonas reinhardtii</i>	PET	[141]
<i>Galleria mellonella</i>	PET	[142]
<i>Eucheuma cottonii</i>	PET	[143]
<i>Chlorella vulgaris</i>	PET	[144]
<i>Dunaliella salina</i>	PET	[145]
<i>Chlorella vulgaris</i>	PHB	[146]
<i>Skeletonema costatum</i>	PET	[147]
<i>Spirulina species</i>	PET	[148]
<i>Arthrospira species</i>	PET	[149]

Biodegradation of MP using worms

PL wastes that have accumulated in the environment are creating an ever-increasing hazard to the ecosystem. Biodegradable plastics are environmentally friendly; they have an expanding range of possible applications, which is fueled by the increased usage of PLs in packaging. For many years, polyethylene was thought to be non-biodegradable. The wax worms that consume plastic were found by chance. Federica Bertocchini of the Institute of Biomedicine and Biotechnology of Cantabria in Spain, a scientist and hobby beekeeper, was irritated when she discovered wax worms in one of her beehives at home. She decided to clean the beehive while going about her business, keeping the wax worms in an ordinary plastic shopping bag. She'd hidden the worms in a plastic bag in another room. When she returned to the room, she was astounded to see worms crawling all over the place. The plastic bag was punctured. This indicated that the wax worms (*Achroia grisella*, *Galleria mellonella* and *Plodia interpunctella*) had bitten their way out of the plastic bags at a fast rate [150]. Other than wax worm's super worms such as *Zophobas atratus* which are used to degrade polystyrene [151] and meal worms of *Enebrio molitor* used for biodegradation of PET and PLs mixtures [152].

CONCLUSION

MP pollution is becoming a growing environmental problem, notably in the world's seas. PET, a polymer utilized in a variety of applications such as textiles and food packaging, is one of the most common components of PL trash. PET is very resistant to environmental biodegradation, resulting in a slew of environmental issues, such organic pollution absorption and concentration, harmful impacts on marine animals, and the spread of potentially invasive species to new habitats. Landfill, incineration, and recycling are the only three large-scale PL disposal technologies now in use. Each method has its own set of downsides and drawbacks. Both landfilling and incineration discharge hazardous secondary pollutants into the environment, and landfilling has the added disadvantage of requiring a considerable amount of land space. Recycling alleviates landfill and incineration-related environmental issues; nonetheless, the process is inefficient, and the deteriorating quality of the PLY generated is a determining factor. Since the process is less cost-effective, there is less incentive for recycling facilities to be built. Biodegradation is a popular alternative for disposing of PL trash in an ecologically acceptable and effective manner. To date, no commercially viable biodegradation protocol for PET has been developed; however, significant research is still being conducted in the field of PLY biodegradation, and given the vast metabolic potential of MO's, it is expected that viable biodegradation processes will be developed in a matter of time.

REFERENCES

- Nicholson, J. L., & Leighton, G. R. (1942). Plastics come of age. *Harper's Magazine*, 185, 300-307.
- Susan Freinkel; *Plastics: A toxic love story* (New York: Henry Holt, 2011), p.4
- Boucher, J., & Friot, D. (2017). Primary microplastics in the oceans: a global evaluation of sources (Vol. 10). Gland, Switzerland: Iucn.
- Suran, M. (2018). A planet too rich in fibre: Microfibre pollution may have major consequences on the environment and human health. *EMBO reports*, 19(9), e46701.
- Wieczorek, A. M., Morrison, L., Croot, P. L., Allcock, A. L., MacLoughlin, E., Savard, O., ... & Doyle, T. K. (2018). Frequency of microplastics in mesopelagic fishes from the Northwest Atlantic. *Frontiers in Marine Science*, 5, 39.
- Thorpe, A., & Harrison, R. M. (2008). Sources and properties of non-exhaust particulate matter from road traffic: a review. *Science of the total environment*, 400(1-3), 270-282.
- Adamiec, E., Jarosz-Krzemińska, E., & Wieszała, R. (2016). Heavy metals from non-exhaust vehicle emissions in urban and motorway road dusts. *Environmental monitoring and assessment*, 188(6), 1-11.
- Amato, F., Karanasiou, A., Moreno, T., Alastuey, A., Orza, J. A. G., Lumberras, J., ... & Querol, X. (2012). Emission factors from road dust resuspension in a Mediterranean freeway. *Atmospheric Environment*, 61, 580-587.
- Bejgarn, S., MacLeod, M., Bogdal, C., & Breitholtz, M. (2015). Toxicity of leachate from weathering plastics: An exploratory screening study with *Nitocra spinipes*. *Chemosphere*, 132, 114-119.
- Chan, D. S. E. A., & Stachowiak, G. W. (2004). Review of automotive brake friction materials. *Proceedings of the Institution of Mechanical Engineers, Part D: Journal of Automobile Engineering*, 218(9), 953-966.
- Malachova, K., Kukutschova, J., Rybkova, Z., Sezimova, H., Placha, D., Cabanova, K., & Filip, P. (2016). Toxicity and mutagenicity of low-metallic automotive brake pad materials. *Ecotoxicology and environmental safety*, 131, 37-44.
- Dahl, A., Gharibi, A., Swietlicki, E., Gudmundsson, A., Bohgard, M., Ljungman, A., ... & Gustafsson, M. (2006). Traffic-generated emissions of ultrafine particles from pavement-tire interface. *Atmospheric Environment*, 40(7), 1314-1323.
- Avagyan, R., Sadiktis, I., Bergvall, C., & Westerholm, R. (2014). Tire tread wear particles in ambient air—a previously unknown source of human exposure to the biocide 2-mercaptobenzothiazole. *Environmental science and pollution research*, 21(19), 11580-11586.
- Kole, P. J., Löhr, A. J., Van Belleghem, F. G., & Ragas, A. M. (2017). Wear and tear of tyres: a stealthy source of microplastics in the environment. *International journal of environmental research and public health*, 14(10), 1265.
- De Falco, F., Cocco, M., Avella, M., & Thompson, R. C. (2020). Microfiber release to water, via laundering, and to air, via everyday use: a comparison between polyester clothing with differing textile parameters. *Environmental science & technology*, 54(6), 3288-3296.
- McIlwraith, H. K., Lin, J., Erdle, L. M., Mallos, N., Diamond, M. L., & Rochman, C. M. (2019). Capturing microfibers—marketed technologies reduce microfiber emissions from washing machines. *Marine pollution bulletin*, 139, 40-45.
- Chan, F., Rajaram, N., House, R., Kudla, I., Lipszyc, J., & Tarlo, S. M. (2017). Potential respiratory effects from 3-D printing. In *B58. Occupational lung disease: case studies, epidemiology, and mechanisms* (pp. A3861-A3861). American Thoracic Society.
- Almroth, B. M. C., Åström, L., Roslund, S., Petersson, H., Johansson, M., & Persson, N. K. (2018). Quantifying shedding of synthetic fibers from textiles; a source of microplastics released into the environment. *Environmental Science and pollution research*, 25(2), 1191-1199.

19. Barlow, C. Y., & Morgan, D. C. (2013). Polymer film packaging for food: An environmental assessment. *Resources, Conservation and Recycling*, 78, 74-80.
20. Brinton, W. I. L. L., Dietz, C. Y. N. D. R. A., Bouyouunan, A. L. Y. C. I. A., & Matsch, D. (2019). The environmental hazards inherent in the composting of plastic-coated paper products. *Eco-Cycle*, (<http://www.ecocycle.org/microplasticsincompost>), Accessed, 3.
21. Hodson, M. E., Duffus-Hodson, C. A., Clark, A., Prendergast-Miller, M. T., & Thorpe, K. L. (2017). Plastic bag derived-microplastics as a vector for metal exposure in terrestrial invertebrates. *Environmental Science & Technology*, 51(8), 4714-4721.
22. Beament, M. (2018). McDonalds to ban plastic straws in all of its restaurants in UK and Ireland. London: Independent Print Limited.
23. Lim, J. W., Ting, D. W. Q., Loh, K. C., Ge, T., & Tong, Y. W. (2018). Effects of disposable plastics and wooden chopsticks on the anaerobic digestion of food waste. *Waste Management*, 79, 607-614.
24. Fendall, L. S., & Sewell, M. A. (2009). Contributing to marine pollution by washing your face: microplastics in facial cleansers. *Marine pollution bulletin*, 58(8), 1225-1228.
25. Cheung, P. K., & Fok, L. (2017). Characterisation of plastic microbeads in facial scrubs and their estimated emissions in Mainland China. *Water research*, 122, 53-61.
26. Anagnosti, L., Varvaresou, A., Pavlou, P., Protopapa, E., & Carayanni, V. (2021). Worldwide actions against plastic pollution from microbeads and microplastics in cosmetics focusing on European policies. Has the issue been handled effectively?. *Marine Pollution Bulletin*, 162, 111883. antifouling paints. *Prog. Org. Coat.*, 59: 2-20.
27. Loretz, L. J., Api, A. M., Babcock, L., Barraj, L. M., Burdick, J., Cater, K. C., ... & Scraftford, C. G. (2008). Exposure data for cosmetic products: facial cleanser, hair conditioner, and eye shadow. *Food and chemical Toxicology*, 46(5), 1516-1524.
28. Boag, A. H., Colby, T. V., Fraire, A. E., Kuhn III, C., Roggli, V. L., Travis, W. D., & Vallyathan, V. (1999). The pathology of interstitial lung disease in nylon flock workers. *The American journal of surgical pathology*, 23(12), 1539.
29. Eschenbacher, W. L., Kreiss, K., Lougheed, M. D., Pransky, G. S., Day, B., & Castellan, R. M. (1999). Nylon flock-associated interstitial lung disease. *American journal of respiratory and critical care medicine*, 159(6), 2003-2008.
30. Warheit, D. B.; Webb, T. R.; Reed, K. L.; Hansen, J. F.; Kennedy, G. L., Jr. Four-week inhalation toxicity study in rats with nylon respirable fibers: rapid lung clearance. *Toxicology* 2003, 192 (2-3), 189-210.
31. Dharmaraj, S., Ashokkumar, V., Hariharan, S., Manibharathi, A., Show, P. L., Chong, C. T., & Ngamcharussrivichai, C. (2021). The COVID-19 pandemic face mask waste: a blooming threat to the marine environment. *Chemosphere*, 272, 129601.
32. Rutala, W. A., & Mayhall, C. G. (1992). Medical waste. *Infection Control & Hospital Epidemiology*, 13(1), 38-48.
33. Joseph, B., James, J., Kalarikkal, N., & Thomas, S. (2021). Recycling of medical plastics. *Advanced Industrial and Engineering Polymer Research*, 4(3), 199-208.
34. Almeida, E., Diamantino, T.C. & de Sousa, O. 2007. Marine paints: the particular case of
35. Antunes, J. C., Frias, J. G. L., Micaelo, A. C., & Sobral, P. (2013). Resin pellets from beaches of the Portuguese coast and adsorbed persistent organic pollutants. *Estuarine, Coastal and Shelf Science*, 130, 62-69.
36. Arnold, S. M., Clark, K. E., Staples, C. A., Klecka, G. M., Dimond, S. S., Caspers, N., & Hentges, S. G. (2013). Relevance of drinking water as a source of human exposure to bisphenol A. *Journal of exposure science & environmental epidemiology*, 23(2), 137-144.
37. Arthur, C., Sutton-Grier, A. E., Murphy, P., & Bamford, H. (2014). Out of sight but not out of mind: harmful effects of derelict traps in selected US coastal waters. *Marine Pollution Bulletin*, 86(1-2), 19-28.
38. Austin, H. M., & Stoops-Glas, P. M. (1977). The distribution of polystyrene spheres and nibs in Block Island Sound during 1972-1973. *Chesapeake Science*, 18(1), 89-92.
39. Bach, C., Dauchy, X., Severin, I., Munoz, J. F., Etienne, S., & Chagnon, M. C. (2013). Effect of temperature on the release of intentionally and non-intentionally added substances from polyethylene terephthalate (PET) bottles into water: Chemical analysis and potential toxicity. *Food Chemistry*, 139(1-4), 672-680.
40. Bains, M., Martellini, T., Cincinelli, A., Campani, T., Minutoli, R., Panti, C., ... & Fossi, M. C. (2017). First detection of seven phthalate esters (PAEs) as plastic tracers in superficial neustonic/planktonic samples and cetacean blubber. *Analytical Methods*, 9(9), 1512-1520.
41. Bernard, L., Vaudaux, P., Huggler, E., Stern, R., Fréhel, C., Francois, P., ... & Hoffmeyer, P. (2007). Inactivation of a subpopulation of human neutrophils by exposure to ultrahigh-molecular-weight polyethylene wear debris. *FEMS Immunology & Medical Microbiology*, 49(3), 425-432.
42. Calafat, A. M., Kuklenyik, Z., Reidy, J. A., Caudill, S. P., Ekong, J., & Needham, L. L. (2005). Urinary concentrations of bisphenol A and 4-nonylphenol in a human reference population.

- Environmental health perspectives, 113(4), 391-395.
43. Cardia, F., & Lovatelli, A. (2015). Aquaculture operations in floating.
44. Zhu, B., Zhu, X., Wang, L., Liang, Y., Feng, Q., & Pan, J. (2017). Functional exploration of the IFT-A complex in intraflagellar transport and ciliogenesis. *PLoS genetics*, 13(2), e1006627.
45. Chua, E. M., Shimeta, J., Nuggeoda, D., Morrison, P. D., & Clarke, B. O. (2014). Assimilation of polybrominated diphenyl ethers from microplastics by the marine amphipod, *Allorchestes compressa*. *Environmental science & technology*, 48(14), 8127-8134.
46. Dantas, D. V., Barletta, M., & Da Costa, M. F. (2012). The seasonal and spatial patterns of ingestion of polyfilament nylon fragments by estuarine drums (*Sciaenidae*). *Environmental Science and Pollution Research*, 19(2), 600-606.
47. Day, R. H., & Shaw, D. G. (1987). Patterns in the abundance of pelagic plastic and tar in the North Pacific Ocean, 1976–1985. *Marine Pollution Bulletin*, 18(6), 311-316.
48. Day, R. H., Shaw, D. G., & Ignell, S. E. (1990). The quantitative distribution and characteristics of neuston plastic in the North Pacific Ocean, 1985–88. In *Proceedings of the 2nd International Conference on Marine Debris* (pp. 247-266).
49. Endo, S., Takizawa, R., Okuda, K., Takada, H., Chiba, K., Kanehiro, H., ... & Date, T. (2005). Concentration of polychlorinated biphenyls (PCBs) in beached resin pellets: variability among individual particles and regional differences. *Marine pollution bulletin*, 50(10), 1103-1114.
50. Gretzer, C., Werthen, M., & Thomsen, P. (2002). Apoptosis and cytokine release in human monocytes cultured on polystyrene and fibrinogen-coated polystyrene surfaces. *Biomaterials*, 23(7), 1639-1648.
51. Harrison, J. P., Schratzberger, M., Sapp, M., & Osborn, A. M. (2014). Rapid bacterial colonization of low-density polyethylene microplastics in coastal sediment microcosms. *BMC microbiology*, 14(1), 1-15.
52. Hillery, A., Jani, P., & Florence, A. (1994). Comparative, quantitative study of lymphoid and non-lymphoid uptake of 60 nm polystyrene particles. *Journal of drug targeting*, 2(2), 151-156.
53. Hu, L., Su, L., Xue, Y., Mu, J., Zhu, J., Xu, J., & Shi, H. (2016). Uptake, accumulation and elimination of polystyrene microspheres in tadpoles of *Xenopus tropicalis*. *Chemosphere*, 164, 611-617.
54. Jang, M., Shim, W. J., Han, G. M., Rani, M., Song, Y. K., & Hong, S. H. (2016). Styrofoam debris as a source of hazardous additives for marine organisms. *Environmental science & technology*, 50(10), 4951-4960.
55. Klärner, P., Klenz, R., Eder, R., Volz, W. E., Schnell, H. W., Leyendecker, D., ... & Christian, M. S. (1998). Preparation and analysis of styrene oligomers containing migrates from various polystyrenes used in food packaging. *Drug and chemical toxicology*, 21(sup1), 31-49.
56. Kukulka, T., Proskurowski, G., Morét-Ferguson, S., Meyer, D. W., & Law, K. L. (2012). The effect of wind mixing on the vertical distribution of buoyant plastic debris. *Geophysical research letters*, 39(7).
57. Mazurais, D., Ernande, B., Quazuguel, P., Severe, A., Huelvan, C., Madec, L., ... & Zambonino-Infante, J. (2015). Evaluation of the impact of polyethylene microbeads ingestion in European sea bass (*Dicentrarchus labrax*) larvae. *Marine environmental research*, 112, 78-85.
58. Pemba, A. G., Rostagno, M., Lee, T. A., & Miller, S. A. (2014). Cyclic and spirocyclic polyacetal ethers from lignin-based aromatics. *Polymer Chemistry*, 5(9), 3214-3221.
59. Ramos, J. A., Barletta, M., & Costa, M. F. (2012). Ingestion of nylon threads by Gerreidae while using a tropical estuary as foraging grounds. *Aquatic Biology*, 17(1), 29-34.
60. Rani, M., Shim, W. J., Han, G. M., Jang, M., Song, Y. K., & Hong, S. H. (2014). Hexabromocyclododecane in polystyrene based consumer products: an evidence of unregulated use. *Chemosphere*, 110, 111-119.
61. Sasamoto, T., Ushio, F., Kikutani, N., Saitoh, Y., Yamaki, Y., Hashimoto, T., ... & Ibe, A. (2006). Estimation of 1999–2004 dietary daily intake of PCDDs, PCDFs and dioxin-like PCBs by a total diet study in metropolitan Tokyo, Japan. *Chemosphere*, 64(4), 634-641.
62. Weinstein, J. E., Crocker, B. K., & Gray, A. D. (2016). From macroplastic to microplastic: Degradation of high-density polyethylene, polypropylene, and polystyrene in a salt marsh habitat. *Environmental Toxicology and Chemistry*, 35(7), 1632-1640.
63. Thomas, S. N., & Hridayanathan, C. (2006). The effect of natural sunlight on the strength of polyamide 6 multifilament and monofilament fishing net materials. *Fisheries research*, 81(2-3), 326-330.
64. Liu, C., Shi, C., Zhu, S., Wei, R., & Yin, C. C. (2019). Structural and functional characterization of polyethylene terephthalate hydrolase from *Ideonella sakaiensis*. *Biochemical and biophysical research communications*, 508(1), 289-294.
65. Hamlin, H. J., Marciano, K., & Downs, C. A. (2015). Migration of nonylphenol from food-grade plastic is toxic to the coral reef fish species *Pseudochromis fridmani*. *Chemosphere*, 139, 223-228.
66. Rist, S. E., Assidqi, K., Zamani, N. P., Appel, D., Perschke, M., Huhn, M., & Lenz, M. (2016). Suspended micro-sized PVC particles impair the performance and decrease survival in the Asian green mussel *Perna viridis*. *Marine pollution bulletin*, 111(1-2), 213-220.

67. Godoy, V., Blázquez, G., Calero, M., Quesada, L., & Martín-Lara, M. A. (2019). The potential of microplastics as carriers of metals. *Environmental Pollution*, 255, 113363.
68. Lomer, M. C., Thompson, R. P., & Powell, J. J. (2002). Fine and ultrafine particles of the diet: influence on the mucosal immune response and association with Crohn's disease. *Proceedings of the Nutrition Society*, 61(1), 123-130.
69. Chiu, H. W., Xia, T., Lee, Y. H., Chen, C. W., Tsai, J. C., & Wang, Y. J. (2015). Cationic polystyrene nanospheres induce autophagic cell death through the induction of endoplasmic reticulum stress. *Nanoscale*, 7(2), 736-746.
70. Lee, D. H. (2018). Evidence of the possible harm of endocrine-disrupting chemicals in humans: ongoing debates and key issues. *Endocrinology and Metabolism*, 33(1), 44-52.
71. Notarstefano, V., Sabbatini, S., Pro, C., Belloni, A., Orilisi, G., Rubini, C., ... & Giorgini, E. (2020). Exploiting fourier transform infrared and Raman microspectroscopies on cancer stem cells from oral squamous cells carcinoma: New evidence of acquired cisplatin chemoresistance. *Analyst*, 145(24), 8038-8049.
72. Schwabl, P., Köppel, S., Königshofer, P., Bucsics, T., Trauner, M., Reiberger, T., & Liebmann, B. (2019). Detection of various microplastics in human stool: a prospective case series. *Annals of internal medicine*, 171(7), 453-457.
73. Kaksonen, M., & Roux, A. (2018). Mechanisms of clathrin-mediated endocytosis. *Nature reviews Molecular cell biology*, 19(5), 313-326.
74. Forte, M., Iachetta, G., Tussellino, M., Carotenuto, R., Prisco, M., De Falco, M., ... & Valiante, S. (2016). Polystyrene nanoparticles internalization in human gastric adenocarcinoma cells. *Toxicology in Vitro*, 31, 126-136.
75. Xia, T., Kovochich, M., Liong, M., Zink, J. I., & Nel, A. E. (2008). Cationic polystyrene nanosphere toxicity depends on cell-specific endocytic and mitochondrial injury pathways. *ACS nano*, 2(1), 85-96.
76. Luo, T., Zhang, Y., Wang, C., Wang, X., Zhou, J., Shen, M., ... & Jin, Y. (2019). Maternal exposure to different sizes of polystyrene microplastics during gestation causes metabolic disorders in their offspring. *Environmental Pollution*, 255, 113122.
77. Hirt, N., & Body-Malapel, M. (2020). Immunotoxicity and intestinal effects of nano-and microplastics: a review of the literature. *Particle and Fibre Toxicology*, 17(1), 1-22.
78. Katznelson, L., Laws Jr, E. R., Melmed, S., Molitch, M. E., Murad, M. H., Utz, A., & Wass, J. A. (2014). Acromegaly: an endocrine society clinical practice guideline. *The Journal of Clinical Endocrinology & Metabolism*, 99(11), 3933-3951.
79. Melzer, D., Rice, N. E., Lewis, C., Henley, W. E., & Galloway, T. S. (2010). Association of urinary bisphenol a concentration with heart disease: evidence from NHANES 2003/06. *PloS one*, 5(1), e8673.
80. Melzer, D., Osborne, N. J., Henley, W. E., Cipelli, R., Young, A., Money, C., ... & Galloway, T. S. (2012). Urinary bisphenol A concentration and risk of future coronary artery disease in apparently healthy men and women. *Circulation*, 125(12), 1482-1490.
81. Brown, D. M., Wilson, M. R., MacNee, W., Stone, V., & Donaldson, K. (2001). Size-dependent proinflammatory effects of ultrafine polystyrene particles: a role for surface area and oxidative stress in the enhanced activity of ultrafines. *Toxicology and applied pharmacology*, 175(3), 191-199.
82. Veruva, S. Y., Lanman, T. H., Isaza, J. E., Freeman, T. A., Kurtz, S. M., & Steinbeck, M. J. (2017). Periprosthetic UHMWPE wear debris induces inflammation, vascularization, and innervation after total disc replacement in the lumbar spine. *Clinical Orthopaedics and Related Research*, 475(5), 1369-1381.
83. Qiao, R., Sheng, C., Lu, Y., Zhang, Y., Ren, H., & Lemos, B. (2019). Microplastics induce intestinal inflammation, oxidative stress, and disorders of metabolome and microbiome in zebrafish. *Science of the Total Environment*, 662, 246-253.
84. de Sá, L. C., Luís, L. G., & Guilhermino, L. (2015). Effects of microplastics on juveniles of the common goby (*Pomatoschistus microps*): confusion with prey, reduction of the predatory performance and efficiency, and possible influence of developmental conditions. *Environmental pollution*, 196, 359-362.
85. Johnson, N. G., Burnett, L. E., & Burnett, K. G. (2011). Properties of bacteria that trigger hemocytopenia in the Atlantic blue crab, *Callinectes sapidus*. *The Biological Bulletin*, 221(2), 164-175.
86. Rochman, C. M., Hoh, E., Kurobe, T., & Teh, S. J. (2013). Ingested plastic transfers hazardous chemicals to fish and induces hepatic stress. *Scientific reports*, 3(1), 1-7.
87. Dawson, A., Huston, W., Kawaguchi, S., King, C., Cropp, R., Wild, S., ... & Bengtson Nash, S. (2018). Uptake and depuration kinetics influence microplastic bioaccumulation and toxicity in Antarctic krill (*Euphausia superba*). *Environmental science & technology*, 52(5), 3195-3201.
88. Rochman, C. M., Kurobe, T., Flores, I., & Teh, S. J. (2014). Early warning signs of endocrine disruption in adult fish from the ingestion of polyethylene with and without sorbed chemical pollutants from the marine environment. *Science of the total environment*, 493, 656-661.
89. Tanaka, K., Takada, H., Yamashita, R., Mizukawa, K., Fukuwaka, M. A., & Watanuki, Y. (2013). Accumulation of plastic-derived chemicals in tissues of seabirds ingesting marine plastics. *Marine pollution bulletin*, 69(1-2), 219-222.

90. Amereh, F., Babaei, M., Eslami, A., Fazelpour, S., & Rafiee, M. (2020). The emerging risk of exposure to nano (micro) plastics on endocrine disturbance and reproductive toxicity: From a hypothetical scenario to a global public health challenge. *Environmental Pollution*, 261, 114158.
91. Ballejo, F., Plaza, P., Speziale, K. L., Lambertucci, A. P., & Lambertucci, S. A. (2021). Plastic ingestion and dispersion by vultures may produce plastic islands in natural areas. *Science of the Total Environment*, 755, 142421.
92. Du, Z. H., Xia, J., Sun, X. C., Li, X. N., Zhang, C., Zhao, H. S., ... & Li, J. L. (2017). A novel nuclear xenobiotic receptors (AhR/PXR/CAR)-mediated mechanism of DEHP-induced cerebellar toxicity in quails (*Coturnix japonica*) via disrupting CYP enzyme system homeostasis. *Environmental Pollution*, 226, 435-443.
93. Li, P. C., Li, X. N., Du, Z. H., Wang, H., Yu, Z. R., & Li, J. L. (2018). Di (2-ethyl hexyl) phthalate (DEHP)-induced kidney injury in quail (*Coturnix japonica*) via inhibiting HSF1/HSF3-dependent heat shock response. *Chemosphere*, 209, 981-988.
94. Omid, A., Naeemipour, H., & Hosseini, M. (2012). Plastic debris in the digestive tract of sheep and goats: an increasing environmental contamination in Birjand, Iran. *Bulletin of environmental contamination and toxicology*, 88(5), 691-694.
95. Wagner, S., Legros, S., Löschner, K., Liu, J., Navratilova, J., Grombe, R., ... & Hofmann, T. (2015). First steps towards a generic sample preparation scheme for inorganic engineered nanoparticles in a complex matrix for detection, characterization, and quantification by asymmetric flow-field flow fractionation coupled to multi-angle light scattering and ICP-MS. *Journal of Analytical Atomic Spectrometry*, 30(6), 1286-1296.
96. Morgado, V., Gomes, L., da Silva, R. B., & Palma, C. (2021). Validated spreadsheet for the identification of PE, PET, PP and PS microplastics by micro-ATR-FTIR spectra with known uncertainty. *Talanta*, 122624.
97. Watteau, F., Dignac, M. F., Bouchard, A., Revallier, A., & Houot, S. (2018). Microplastic detection in soil amended with municipal solid waste composts as revealed by transmission electronic microscopy and pyrolysis/GC/MS. *Frontiers in Sustainable Food Systems*, 2, 81.
98. Liu, Y., Li, R., Yu, J., Ni, F., Sheng, Y., Scircle, A., ... & Zhou, Y. (2021). Separation and identification of microplastics in marine organisms by TGA-FTIR-GC/MS: A case study of mussels from coastal China. *Environmental Pollution*, 272, 115946.
99. Hendrickson, E., Minor, E. C., & Schreiner, K. (2018). Microplastic abundance and composition in western Lake Superior as determined via microscopy, Pyr-GC/MS, and FTIR. *Environmental science & technology*, 52(4), 1787-1796.
100. Olesen, K. B., van Alst, N., Simon, M., Vianello, A., Liu, F., & Vollertsen, J. (2017). Analysis of microplastics using FTIR imaging: application note. Agilent Application Note Environment.
101. Müsellim, E., Tahir, M. H., Ahmad, M. S., & Ceylan, S. (2018). Thermokinetic and TG/DSC-FTIR study of pea waste biomass pyrolysis. *Applied Thermal Engineering*, 137, 54-61.
102. Bitter, H., & Lackner, S. (2021). Fast and easy quantification of semi-crystalline microplastics in exemplary environmental matrices by differential scanning calorimetry (DSC). *Chemical Engineering Journal*, 423, 129941.
103. Majewsky, M., Bitter, H., Eiche, E., & Horn, H. (2016). Determination of microplastic polyethylene (PE) and polypropylene (PP) in environmental samples using thermal analysis (TGA-DSC). *Science of the Total Environment*, 568, 507-511.
104. Luo, H., Xiang, Y., Li, Y., Zhao, Y., & Pan, X. (2021). Photocatalytic aging process of Nano-TiO₂ coated polypropylene microplastics: Combining atomic force microscopy and infrared spectroscopy (AFM-IR) for nanoscale chemical characterization. *Journal of Hazardous Materials*, 404, 124159.
105. Abbas-Abadi, M. S. (2021). The effect of process and structural parameters on the stability, thermo-mechanical and thermal degradation of polymers with hydrocarbon skeleton containing PE, PP, PS, PVC, NR, PBR and SBR. *Journal of Thermal Analysis and Calorimetry*, 143(4), 2867-2882.
106. Fernandes, E. G., Kenawy, E. R., Miertus, S., & Chiellini, E. (2002). Environmentally degradable plastics: thermal behavior of polymer blends based on waste gelatin. *Polimery*, 47(7-8), 500-508.
107. Santos, A. S. F., Agnelli, J. A. M., Trevisan, D. W., & Manrich, S. (2002). Degradation and stabilization of polyolefins from municipal plastic waste during multiple extrusions under different reprocessing conditions. *Polymer Degradation and Stability*, 77(3), 441-447.
108. Raghavan, D. (1995). Characterization of biodegradable plastics. *Polymer-Plastics Technology and Engineering*, 34(1), 41-63.
109. Shah, T. V., & Vasava, D. V. (2019). A glimpse of biodegradable polymers and their biomedical applications. *e-Polymers*, 19(1), 385-410.
110. Karamanlioglu, M., Preziosi, R., and Robson, G. D. (2017). Abiotic and biotic environmental degradation of the bioplastic polymer poly(lactic acid): a review. *Polym. Degrad. Stab.* 137, 122-130.
111. Restrepo-Flórez, J. M., Bassi, A., & Thompson, M. R. (2014). Microbial degradation and deterioration of polyethylene—A review. *International Biodeterioration & Biodegradation*, 88, 83-90.
112. Huang, X., Cao, L., Qin, Z., Li, S., Kong, W., & Liu, Y. (2018). Tat-independent secretion of polyethylene terephthalate hydrolase PETase in

- Bacillus subtilis* 168 mediated by its native signal peptide. *Journal of agricultural and food chemistry*, 66(50), 13217-13227.
113. Seo, H., Kim, S., Son, H. F., Sagong, H. Y., Joo, S., & Kim, K. J. (2019). Production of extracellular PETase from *Ideonella sakaiensis* using sec-dependent signal peptides in *E. coli*. *Biochemical and biophysical research communications*, 508(1), 250-255.
 114. Tanasupawat, S., Takehana, T., Yoshida, S., Hiraga, K., & Oda, K. (2016). *Ideonella sakaiensis* sp. nov., isolated from a microbial consortium that degrades poly (ethylene terephthalate). *International journal of systematic and evolutionary microbiology*, 66(8), 2813-2818.
 115. Fecker, T., Galaz-Davison, P., Engelberger, F., Narui, Y., Sotomayor, M., Parra, L. P., & Ramírez-Sarmiento, C. A. (2018). Active site flexibility as a hallmark for efficient PET degradation by *I. sakaiensis* PETase. *Biophysical journal*, 114(6), 1302-1312.
 116. Khoironi, A., & Anggoro, S. (2019). Evaluation of the interaction among microalgae *Spirulina* sp, plastics polyethylene terephthalate and polypropylene in freshwater environment. *Journal of Ecological Engineering*, 20(6).
 117. Chavant, P., Martinie, B., Meylheuc, T., Bellon-Fontaine, M. N., & Hebraud, M. (2002). *Listeria monocytogenes* LO28: surface physicochemical properties and ability to form biofilms at different temperatures and growth phases. *Applied and environmental microbiology*, 68(2), 728-737.
 118. Yamada-Onodera, K., Mukumoto, H., Katsuyaya, Y., Saiganji, A., & Tani, Y. (2001). Degradation of polyethylene by a fungus, *Penicillium simplicissimum* YK. *Polymer degradation and stability*, 72(2), 323-327.
 119. Hadad, D., Geresh, S., & Sivan, A. (2005). Biodegradation of polyethylene by the thermophilic bacterium *Brevibacillus borstelensis*. *Journal of applied microbiology*, 98(5), 1093-1100.
 120. Pometto, A. L., Lee, B. T., & Johnson, K. E. (1992). Production of an extracellular polyethylene-degrading enzyme (s) by *Streptomyces* species. *Applied and environmental microbiology*, 58(2), 731-733.
 121. Mabrouk, M. M., & Sabry, S. A. (2001). Degradation of poly (3-hydroxybutyrate) and its copolymer poly (3-hydroxybutyrate-co-3-hydroxyvalerate) by a marine *Streptomyces* sp. SNG9. *Microbiological research*, 156(4), 323-335.
 122. Kita, K., Ishimaru, K., Teraoka, M., Yanase, H., & Kato, N. (1995). Properties of poly (3-hydroxybutyrate) depolymerase from a marine bacterium, *Alcaligenes faecalis* AE122. *Applied and Environmental Microbiology*, 61(5), 1727-1730.
 123. Oda, Y., Oida, N., Urakami, T., & Tonomura, K. (1997). Polycaprolactone depolymerase produced by the bacterium *Alcaligenes faecalis*. *FEMS Microbiology letters*, 152(2), 339-343.
 124. Akutsu, Y., Nakajima-Kambe, T., Nomura, N., & Nakahara, T. (1998). Purification and properties of a polyester polyurethane-degrading enzyme from *Comamonas acidovorans* TB-35. *Applied and Environmental Microbiology*, 64(1), 62-67.
 125. Ali, M. I., Ahmed, S., Javed, I., Ali, N., Atiq, N., Hameed, A., & Robson, G. (2014). Biodegradation of starch blended polyvinyl chloride films by isolated *Phanerochaete chrysosporium* PV1. *International Journal of Environmental Science and Technology*, 11(2), 339-348.
 126. Creamer, N. J., Baxter-Plant, V. S., Henderson, J., Potter, M., & Macaskie, L. E. (2006). Palladium and gold removal and recovery from precious metal solutions and electronic scrap leachates by *Desulfovibrio desulfuricans*. *Biotechnology letters*, 28(18), 1475-1484.
 127. Allen, A. B., Hilliard, N. P., & Howard, G. T. (1999). Purification and characterization of a soluble polyurethane degrading enzyme from *Comamonas acidovorans*. *International biodeterioration & biodegradation*, 43(1-2), 37-41.
 128. Arias, S., Olivera, E. R., Arcos, M., Naharro, G., & Luengo, J. M. (2008). Genetic analyses and molecular characterization of the pathways involved in the conversion of 2-phenylethylamine and 2-phenylethanol into phenylacetic acid in *Pseudomonas putida* U. *Environmental microbiology*, 10(2), 413-432.
 129. Arkatkar, A., Juwarkar, A. A., Bhaduri, S., Uppara, P. V., & Doble, M. (2010). Growth of *Pseudomonas* and *Bacillus* biofilms on pretreated polypropylene surface. *International Biodeterioration & Biodegradation*, 64(6), 530-536.
 130. Manderfield, L. J., & George Jr, A. L. (2008). KCNE4 can co-associate with the IKs (KCNQ1-KCNE1) channel complex. *The FEBS journal*, 275(6), 1336-1349.
 131. Das, G., Bordoloi, N. K., Rai, S. K., Mukherjee, A. K., & Karak, N. (2012). Biodegradable and biocompatible epoxidized vegetable oil modified thermostable poly (vinyl chloride): Thermal and performance characteristics post biodegradation with *Pseudomonas aeruginosa* and *Achromobacter* sp. *Journal of hazardous materials*, 209, 434-442.
 132. Yin, C. F., Xu, Y., & Zhou, N. Y. (2020). Biodegradation of polyethylene mulching films by a co-culture of *Acinetobacter* sp. strain NyZ450 and *Bacillus* sp. strain NyZ451 isolated from *Tenebrio molitor* larvae. *International Biodeterioration & Biodegradation*, 155, 105089.
 133. Sharma, M., Saini, G. S., Prashar, E., Aggarwal, R., & Kaur, G. (2014). The Effect of World Markets on Indian Policy Rates. *The Journal of Wealth Management*, 16(4), 43-47.
 134. Sarmah, P., & Rout, J. (2018). Efficient biodegradation of low-density polyethylene by cyanobacteria isolated from submerged

- polyethylene surface in domestic sewage water. *Environmental Science and Pollution Research*, 25(33), 33508-33520.
135. Chinaglia, S., Tosin, M., & Degli-Innocenti, F. (2018). Biodegradation rate of biodegradable plastics at molecular level. *Polymer Degradation and Stability*, 147, 237-244.
136. Kumar, R. V., Kanna, G. R., & Elumalai, S. (2017). Biodegradation of polyethylene by green photosynthetic microalgae. *J Bioremediat Biodegrad*, 8(381), 2.
137. Chia, W. Y., Tang, D. Y. Y., Khoo, K. S., Lup, A. N. K., & Chew, K. W. (2020). Nature's fight against plastic pollution: Algae for plastic biodegradation and bioplastics production. *Environmental Science and Ecotechnology*, 100065.
138. Sanniyasi, E., Gopal, R. K., Gunasekar, D. K., & Raj, P. P. (2021). Biodegradation of low-density polyethylene (LDPE) sheet by microalga, *Uronema africanum* Borge. *Scientific Reports*, 11(1), 1-33.
139. Bhuyar, P., Sundararaju, S., Feng, H. X., Rahim, M. H. A., Muniyasamy, S., Maniam, G. P., & Govindan, N. (2021). Evaluation of Microalgae's Plastic Biodeterioration Property by a Consortium of *Chlorella* sp. and *Cyanobacteria* sp. *Environmental Research, Engineering and Management*, 77(3), 86-98.
140. Kadar, N. A. H. A., Rahim, N. S., Yusof, R., Nasir, N. A. H. A., & Hamid, H. A. (2021). A REVIEW ON POTENTIAL OF ALGAE IN PRODUCING BIODEGRADABLE PLASTIC. *International Journal of Engineering Advanced Research*, 3(1), 13-26.
141. Kim, J. W., Park, S. B., Tran, Q. G., Cho, D. H., Choi, D. Y., Lee, Y. J., & Kim, H. S. (2020). Functional expression of polyethylene terephthalate-degrading enzyme (PETase) in green microalgae. *Microbial cell factories*, 19(1), 1-9.
142. Kong, H. G., Kim, H. H., Chung, J. H., Jun, J., Lee, S., Kim, H. M., ... & Ryu, C. M. (2019). The *Galleria mellonella* hologenome supports microbiota-independent metabolism of long-chain hydrocarbon beeswax. *Cell reports*, 26(9), 2451-2464.
143. Consebit, K. L., Dermil, K. C., Magbanua, E. Y., Racadio, F. J., Saavedra, S. V., Abusama, H., & Valdez, A. (2022). Bioplastic from Seaweeds (*Eucheuma Cottonii*) as an Alternative Plastic. *ASEAN Journal of Science and Engineering*, 2(2), 129-132.
144. Falah, W., CHEN, F., Zeb, B. S., Hayat, M. T., Mahmood, Q., Ebadi, A., ... & Li, E. Z. (2020). Polyethylene Terephthalate Degradation by Microalga *Chlorella vulgaris* Along with Pretreatment. *Mater. Plastics*, 57(3), 260-270.
145. Mishra, A., Kavita, K., & Jha, B. (2011). Characterization of extracellular polymeric substances produced by micro-algae *Dunaliella salina*. *Carbohydrate Polymers*, 83(2), 852-857.
146. Robert, R., & Iyer, P. R. (2018). Isolation and Optimization of PHB (Polyhydroxybutyrate) Based Biodegradable Plastics from *Chlorella vulgaris*. *Journal of Bioremediation and Biodegradation*, 1-4.
147. Vickers, N. J. (2017). Animal communication: when i'm calling you, will you answer too?. *Current biology*, 27(14), R713-R715.
148. Dianratri, I., Hadiyanto, H., Khoironi, A., & Pratiwi, W. Z. (2020). The influence of polypropylene and polyethylene microplastics on the quality of spirulina sp. Harvests. *Food Res*, 4, 1739-1743.
149. Chentir, I., Hamdi, M., Doumandji, A., HadjSadok, A., Ouada, H. B., Nasri, M., & Jridi, M. (2017). Enhancement of extracellular polymeric substances (EPS) production in *Spirulina* (*Arthrospira* sp.) by two-step cultivation process and partial characterization of their polysaccharidic moiety. *International journal of biological macromolecules*, 105, 1412-1420.
150. Khyade, V. B. (2018). Review On Biodegradation of Plastic Through Waxworm (Order: Lepidoptera; Family: Pyralidae). *International Academic Journal of Economics*, 5(4), 31-38.
151. Yang, Y., Wang, J., & Xia, M. (2020). Biodegradation and mineralization of polystyrene by plastic-eating superworms *Zophobas atratus*. *Science of the total environment*, 708, 135233.
152. Brandon, A. M., Gao, S. H., Tian, R., Ning, D., Yang, S. S., Zhou, J., ... & Criddle, C. S. (2018). Biodegradation of polyethylene and plastic mixtures in mealworms (larvae of *Tenebrio molitor*) and effects on the gut microbiome. *Environmental science & technology*, 52(11), 6526-6533.